


Microscale Transport Processes
Prof. S.Ganguly
Department of Chemical Engineering
Indian Institute of Technology, Kharagpur

Lecture No.#03
Lab on Chip

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Components of lab on chip

- Pumping
 - Centrifugal force
 - Surface force
 - Electrokinetic force
 - mechanical
- Valve
 - Hydrogel
 - Hydrophobic layer
 - Mechanical
- Separation
 - Field flow fractionation (electrical, thermal, flow)
 - Electrophoresis, Dielectrophoresis, DEP + FFF
 - Diffusion based separation (H-Filter)




I welcome you all to this lecture on Microscale transport process. Here we are going to discuss, various components of lab on chip. As we mentioned in the last class that there are several components we must have in lab on chip device.

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Components of microfluidic device ..contd.

- Mixing
 - Passive using grooves, laminations
 - Active
- Diffusion between layers
 - T-Sensor
- Heating
 - Cyclic heating for PCR reaction – DNA hybridization
- Detection
 - Optical interrogation
 - Amperometric sensing

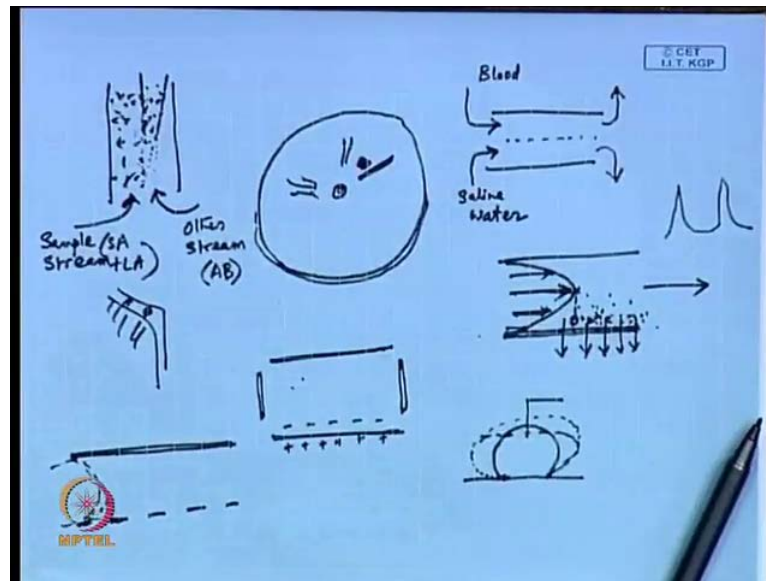
Flow is laminar and the interaction between layers are utilized in most of these components



These components are supposed to be pumping, valve, separation, mixing, diffusion between layers, heating and detection; and there could be some other components also depending on the application that you are looking into. And in all these components, one essential feature is that it is assumed flow is laminar. In fact, flow is very much, in fact the Reynolds number is pretty small in flows in microchannels. Flow is considered laminar, and the interaction between layers are utilized in most of these components. So, this is unlike a typical flow in a channel where there could be turbulence, and there could be movement of ADs from one place to another; that is absolutely missing, because you are relying on two layers flowing like a pack of cards, one sliding against the other. And diffusion is taking place from one layer to the other. So, you are relying on this laminar flow that is for certain. So, these are some of things that you should remember.

Now, when it comes to pumping, of course you can have a pump that we have in a conventional sense; for example, you have seen centrifugal pumps, where you have an impeller rotating. Now, having such components in microscale is not something very easy. Of course, it is durable, it people have done it, but it is not very easy to implement. So, people have looked for other alternative routes that they can use to implement pumping action. And the action that they have is I have categorized them, one is centrifugal force.

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How does centrifugal force work? Suppose, you have a CD type device; suppose you have a CD device, and in this device you have this sample that is going in that sample goes in through the center. So, you have a CD type device, which can be put on a turn table. And it can rotate. Now and it, so it is a plate in which you have curved channels in radial directions in various, several channels in radial directions; and in some of the places, you have reagents stacked reagents put there and you introduce the sample at the center. Then what do you do? You rotate the turn table. So, as you rotate the turn table, because of centrifugal action the fluid will start moving towards the periphery. So, this is an action that you can have this is a, this an action that you can have for pumping fluids. So, you have a CD type device on which you have channels curved and on top of that you have a lead and everything is happening in microscale.

Put the CD device on a turn table, if you introduce the sample at the center of the CD and rotate the CD, automatically the fluid will move towards the periphery. So, this is a centrifugal action that you can rely on for pumping fluids, because you can control the spinning rate and you can control the RPM; and by that you can control at what velocity the sample would be flowing towards the periphery. So, this can be a device. Similarly, you can have surface force. How that force look like? You have all studied capillary rise; if you dip a glass capillary inside water there would be rise of water through the capillary.

How does it happen? The liquid has a weight, but that weight is balanced by surface tension force, because glass is hydrophilic; so you have, if the surface tension force is balanced, the weight of the liquid is balanced by the surface tension force. So, surface tension can raise water through certain height. Now, if you have a tapping point, because anyway you are working with small scales, if you have a tapping point, little above the water level, you can have a continuous flow of water. So, this is basically a surface using surface force to pump a fluid. Next, you can have what you can have is Electrokinetic force. Electrokinetic force means, you can have electrodes put there and you can mobilize the charges. And with the charge, some bulk liquid is also dragged; we will discuss this electrokinetic force in details later. And mechanical of course, you can have actual movement, I mean something similar to centrifugal pump or reciprocating pump, you can have a moving element inside the microchannel that is also a possibility. But other elements are very much perused; other elements are very much looked into for pumping fluid.

The next item is valve. Of course, you can have valve in a conventional, the way you have in a conventional sense, like you have a valve seat, you have a closer of the valve, you have another element that goes in and sticks to the valve seat and so you can consider the valve to be closed. So, you can have such elements design such elements implemented in microscale, but that is not again very easy. However, other way is people have looked into and one is having a Hydrogel layer, because Hydrogel have some unique properties, it can respond to pH or thermal trigger. Either it can contract; I mean it can contract as well as it can expand depending upon the external trigger.

So, if you put a Hydrogel pillar at the channel inlet, and if you put a trigger from outside and by the trigger, you can shrink the gel layer or you can expand gel layer. So by that way, you can block a channel or open part of the channel. So, Hydrogel can act as a valve in that case. Also there could be something called hydrophobic layer. If you have a hydrophobic element on the wall, you know that the contact angle; if there would be adverse contact angle. You might have seen already, when we dip glass capillary in mercury, how the interface would be; so that gives you an example, how hydrophobic layer would have how hydrophobic surface would be acting towards water.

So, if you have an aqua solution and if you put a hydrophobic layer, then because of this surface act, because of this surface tension and contact angle, you may have to cross a threshold pressure, and then only you can have a flow through the channel. So a hydrophobic layer, small hydrophobic layer put on the channel valve can act as a valve in a sense. The third element that we have here is separation in lab on chip device; you need to have separation accomplished. And for that there are various techniques that are available. First, I would you like to talk about is called what is called field flow fractionation. It can be of various types, for example, electrical, thermal or flow. The field flow fractionation, the way it works is, suppose for example, I look at say flow field flow fractionation of flow type. Suppose, you have a channel and on the wall, you have a membrane placed **on the wall you have a membrane placed**. I say that the flow would be laminar, so the velocity profile would be parabolic like this.

And there is this membrane, so if you have solutes inside this, I mean the liquid that is flowing within the channel if that has solutes, then the solutes would be as you collect fluid from this side I mean. So, you have a flow down this way and at the same time, you have some collection of fluid, at a very low rate you are collecting fluids from this side. So, when you are collecting fluid collecting the solvent through this side and you are having solute been deposited, what you eventually have is something called concentration polarization. By that what I mean is you will see that the particles that are being held on the membrane surface, if the particle is smaller, it will have a larger diffusivity and it will be easy for that particle to go to the bulk, move to the bulk.

The particle that is being restrained by the membrane, if it is small, it can go to the bulk easily; if it is a bigger particle, it would be difficult for that particle to go to the bulk. So depending on the size of the particle, you will see a classification here, this is something called a polarization; you will have particles of certain sizes held next to the membrane and particles of other sizes are more close to the bulk, I mean more towards the center. Now, you are introducing a parabolic velocity profile from this side; so that means, this layer is moving faster than the other, this layer is moving little slower and this layer next to the moving at the slowest at smallest velocity. So, if you put something called a fractogram, if you analyze the effluent coming out of this, you will find peaks coming like this. And each peak represents particles of certain size. So, you can classify the particles.

Now suppose, you if you have this device already calibrated with a known mixture of particles and then, if you introduce this if you introduce an unknown sample into this device; and output that you get, if you look it look at it (()) the one that you have the calibration that you have, you can figure out what would be the what is the what are the various components that are; what are the various sizes that are therein this fluid. So, this separation this can be this is referred as field flow fractionation of flow type. Now, here the flow has been put as a transverse gradient. Now, for other types of field flow fractionation, for example, the thermal one, there you have a thermal gradient in the transverse directions. Or electrical field flow fractionation, there you have this electrical gradient transverse to the flow direction. So by that way, you can **by that way you can classified** by that way you can separate particles depending on their sizes or depending upon their properties with a distance from the wall.

And since you have always a parabolic velocity profile, I categorically said that you are relying on laminar flow in the microchannel. So, you can get layers one layer at a time, middle layer first, because that is moving at a highest velocity, next layer and then the other layer. So, you can separate the particles you can get the fractogram with particles arrive eluting at the outlet at a different times. So, you can classify them. Similarly, there are other separation components, like you have this electrophoresis, Dielectrophoresis or Dielectrophoresis plus field flow fractionation. We will discuss this in details in electrophoresis, you have put electrodes and you apply electric field depending on the size and **mobility**, depending on the mobility of the ions; that means, size and charge of the ions, you can separate they can put it will be more closed to the electrode or away from the electro, depending on the mobility of that ion mobility of the charge. So, you can find out what then if you study them.

For example, in electrical field flow fractionation, I will discuss this later that you impose this electric field in transverse direction. So, depending on the mobility of the ions, they will be located either next to the electrode or away from the electrode at various locations. And since you have the laminar flow taking one layer and the next layer, so you will be collecting the particles at different times; and so, you can always using a calibration, you can always figure out particles of what sizes, where there or what mobility where there in the original sample.

Then there is this diffusion best separation, which is referred as H-filter; it depends totally diffusion is does not have any electric field in water. I will discuss this; I will show draw a picture very soon. So, these are the some of the separation components. So, we if you think of conventional chemical applications, there are separation, possible, but when it comes to microscale, you are relying more on this laminar flow and relying more on this parabolic velocity profile and some diffusion or certain responds to electric fields, so these are the some of the things that you look into. Of course, you need mixing in microchannel and mixing is in a traditional sense in a microscale, it is done using an impeller, using a stirrer. Here that is possible, you can have element within a microchannel, but implementing it in microscale is not very easy. So, there are alternative routes people have looked into, one is passive mixing device using grooves or laminations.

You have already heard of these passive mixers that mean, you have to strength flowing through a tube and then you have several baffles placed on the way, so that two streams they come together and then they divert. So by that way, you increase the contact area between the two streams, and so you enhance the mixing. So, these are some of the elements that you would look into, when it comes to mixing in microscale. Then there is diffusion between layers, which is very important, I will discuss this very soon what is a T-Sensor? In T-Sensor diffusion people relying on this laminar flow parabolic or one layer sliding against the other, that means no cross flow no deformation; and then the diffusion from one layer to other this is people relying on this, to develop sensors. So this is a T-Sensor we are going to discuss very soon. Then there is this heating component also you should have in microscale, microscale process. And heating component one heating requirement, I have given an example here cycling cyclic heating for PCR reaction.

These reaction this requires heating in steps, within a very short time we have to accomplish several up and down in temperature. That means, you increase the temperature by certain degrees and then you decrease it, again you increase, again you decrease, again you increase, again you decrease. So, you have to do these are called thermal cycles and you have to expose the sample to such thermal cycles several such cycles within a very short time.

And typically, if you are if the mass is more, then accomplishing these cycles will not be very easy, because if the mass is more you think of it. What would be the heat


requirement, what would be Δh ? Heat requirement would be mass into specific heat into Δt . So, if Δt is known, specific it is known, so, if you have a large thermal mass. So, you have to inject so much of heat and that heat has to distribute through the entire material; and then the temperature has to uniformly rise everywhere within this material. And again you are cooling it down and you are doing it twenty times within a very short period. So that is very effectively done in a microscale and that is what I referred as cyclic heating for PCR reaction.

There are of course, all these after all these microfluidic elements microchannel devices a major application is in sensors. So, there has to be some detection element involved. This detection is done either by optical method, optical interrogation or amperometric sensing; that means, you find out the conductivity of the fluid that is passing by. Say for example, you are you have classified the sample classified, you have a sample and you have classified it into several say several components. And you had this parabolic velocity profile, so one component is eluting at some time, then the next component, then the next component it comes like this. Now, if you want to identify those components, one could be this optical interrogation, optical detection. The other way to do it is by amperometric sensing; that means, you find out what is the conductivity of the sample that is coming out at a particular time, from the outlet and from that you can generate a fractogram. So, these are some of the elements that you have for lab on a chip device.

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Pumping

- Use of moving parts as in a conventional pump with the help of micromachining.
- Centrifugal force to drive fluid through channels in radial direction (lab on CD).
- Use of coating with favorable contact angle, and pillars in the channel to enhance a "capillary rise" type flow.
- Electro-osmosis: Polar liquid in contact with solid wall induces surface charges, which in turn influences migration of charges within the liquid near the wall. Voltage gradient along the length of the channel pulls the charges, and the bulk liquid along with it.
- Electro-wetting: The change in contact angle of a droplet on a surface when an electric field is present at an interface. A droplet is held between two sets of planar electrodes (upper one consists of single continuous ground electrode, and bottom one with an array of independently addressable control electrodes). By spreading the droplet using the electric field such that droplet touches adjacent electrode in the array, and then switching on the adjacent electrode movement of droplet is accomplished.



NPTEL

Now, let us look to this quickly this, what are the pumping elements that we have here. First of all, use of moving parts as in a conventional pump with the help of micromachining this can be done. There can be a moving part, but this can this is not the only way to do and this may not be very easy to accomplish; so, there are other methods which are in demand here. Centrifugal force drive fluids through channels in radial direction, which is referred as lab on CD, this is used in certain industry these lab on CD is used. So, these are I already I have discussed here. The third one is use of coating with favorable contact angle and pillars in the channel to enhance a capillary rise type flow. So, you can in a channel, you can have favorable contact angle as I said capillary rise of water in a glass capillary.

Now, one thing is there the capillary diameter has to be, I think what we have studied is it should be less than say 5 millimeter or there was threshold, I mean, if we cross the diameter, you see these capillary rise is not substantial. So you can, so what you can have is to induce these capillary rise, to induce this surface tension driven flow, you can introduce small pillars on in the channel. So that means if this is the channel, within the channel you can introduce small pillars. So this pillars will provide extra surface area, so that you can have enhanced flow. The third, the fourth element what you have here is electro-osmosis, I said electrokinetic pumping. So electro-osmosis, it is basically like this; a polar liquid in contact with solid wall induces surface charges, which in turn influences migration of charges within the liquid near the wall.

Voltage gradient along the length of the channel of pulls the charges and the bulk liquid along with it. What you have is, you have a wall and you have electrodes placed here. Now, polar liquid in contact with solid wall induces surface charges. So surface is where? This is the surface; this is the wall, so this is the surface. On the surface, there are some surface charges generated. Say I say I write plus, we will discuss this later, we will solve the, we will get in to the theories of it. If you have certain charges induced, so automatically the... So, this polar liquid in contact is solid wall induces surface charges. So, the negative charges within the liquid, they will start getting accumulated on the wall. And then this electrode, they will be pulling these negative charges say. While pulling it, it will pull the bulk liquid along with it. So that is that is how we generate some flow. Of course, the channel this wall should be very close to each other, I mean if you

have a large pipe and if you expect this kind of flow to take place that may not be very much possible or feasible.

The other element that we have is electro-wetting. Here, the change in contact angle of a droplet on a surface, when an electric field is present at an interface. I have supposed the wall and on which I have a droplet sitting there; this is a droplet. Now, if I introduce some voltage difference, if I introduce an electric field, I will find that this contact angle changes, this contact angle changes, if I introduce an electric field there. So, if this contact angle changes, so then this would be more spreading. For example, this was the liquid and later on, when you bring in the electric field, the liquid cannot grow, but the volume remains the same, but the liquid's contact angle will be different. So, if you have an arrangement, where you have suppose, one ground electrode on top and several control electrodes in the bottom.

And suppose, you have a droplet sitting here, a droplet is there and you apply electric field using the electrodes. So, this would more spreading and by while spray while this droplet spreads it will go to the next, it will touch the next electrode. Then what you do is you switch off the, you have these electrodes they can be control separately. So you switch off this electrode and introduce electric field through this electrode. So automatically this droplet will be picked up by this electrode and the same process will continue. So, you will see this droplet rolling down, this droplet will be rolling down through this electrode assembly by this (()).

So that is what exactly what I mean by this electro-wetting here. The change in contact angle of droplet on a surface, when you an electric field is present at an interface. A droplet is held between two sets of planer electrodes, I described here; two sets of planer electrodes, upper one consist of single continues ground electrode and the bottom one, with an array of independently addressable control electrodes. By spreading the droplet using the electric field, such that droplet touches adjacent electrode in the array and then switching on the adjacent electrode, movement of droplet is accomplished. So, this is one method by which you can move droplet within a microchannel.


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T-sensor

- **Sample Stream:**
 - Sample antigen to be measured (SA)
 - Fluorescently labeled antigen (LA) kept to a concentration 2-3 orders of magnitude less than anticipated SA concentration
- **Other Stream:**
 - Known concentration of antibody (AB) to the target analyte

AB molecules are larger and slow to diffuse

1. AB binds with all LA if SA is not target analyte → Bound LA cannot diffuse into AB stream → color stays near interface
2. AB binds with SA and some LA if SA is target analyte → Lot of free LA and free SA to diffuse into AB stream → spread of color into AB stream.
3. More than one T-sensors in the chip with different AB to target analyte.

 Spread of color can be determined digitally.

Next, what we want to discuss is at T-Sensor, because this is some element which you have probably not seen in a microscale. This is a unique device, but this has become once upon a time very popular in microfluidic literature. This T-Sensor, it is basically nothing but a channel, it is nothing but a channel through which two, there is provision for two liquids to enter into the channel, two liquids can enter into the channel. So ideally you expect the liquid to be moving like this, parallelly, like two layers the two layers they are moving parallelly. So, this is ideally that is what you expect, this is a channel with two liquids introduced. So, they will be moving parallelly, one layer sliding against the other. Now if you have, if you allow diffusion to take place in between, then you expect, suppose one is say having red color and other is colorless and one can mix with the other.

So in that case, you can have the color moving from one layer to the other. So, if the red color is, if this is say this is red color and this is say black this is a black color, you will find there would be some amount of diffusion. So, the interface would be more interface would be little blurred, because one layer will diffuse into the other. Now, what you have in a T-Sensor here is that you have a sample stream and other stream. So one is your, one is sample stream and other the other stream I am calling it say other stream. Now, sample stream contains sample antigen that you want to measure, and also a fluorescently labeled antigen that is kept to a concentration two to three orders of magnitude less than anticipated sample antigen concentration.

So, you have a sample stream that contains sample antigen and a fluorescently labeled antigen, which we refer as LA. So, there is SA and there is this LA. So, one stream that is the sample stream, this sample stream contains SA and LA. So, SA plus LA that is what sample stream contains. And this other stream, that contains known concentration of antibody AB to the target analyte. So what that means is I want to find out what pathogen is there in a sample. So, I have several antibodies present and I would like to find out which antibody goes with the sample. So, I say I have say five antibodies and I know four will not go with the sample, the only one will go with the sample that is that target; that but that I do not know, I do not know what is this sample and antigen. So, sample antigen is something which we do which I do not know and I want to find out, what is the sample antigen.

So, I have introduced some amount of fluorescently labeled antigen that is LA, which I introduced with the sample stream, but the LA concentration is very small. And then, I am having this two streams flowing side by side through this channel. So, the implication of this is that AB molecules, these antibody molecules they are larger and slow to diffuse. Now, AB binds with all LA, if SA is not target analyte. See AB binds with LA; AB binds with SA, only if that is the corresponding AB. Because, I have five such AB's present, five such antibodies present, out of them only one can bind with the bind with SA, other will not. So, AB binds with all LA, if SA is not target analyte. So, bound LA cannot diffuse into AB stream; once this LA is bound with the AB, because AB is a bulk here. So, color stays near interface, because LA gives you the color. LA gives you the color, so LA so, color stays near interface, because LA which gives the color that is bound with the bound with AB and AB is a bulkier molecules, so it cannot now diffuse.

So, it will stay next to the interface. AB binds with SA and some LA, if SA is target analyte. So, if SA is target analyte, because SA is there in very large quantity; at the very outside I said that LA concentration is intentionally kept two to three orders of magnitude less than anticipated SA concentration. So, if in the event SA is the target analyte, if in the event AB binds with this analyte; so in that case, there will be lot of free LA available. And this and free SA as well, because AB is all in fix quality and if this is binding both with LA and with SA. So there will be free SA, free LA available and this can diffuse into AB stream.

So, AB stream means which stream, the other stream. So, I had originally sample which is SA plus LA and I have another stream, which is AB. now two are moving side by side and then I will find that in most of the cases, there would be say this is the color. So, these color is penetrating maybe this is more deep and this is light. So, this part is probably light lighter, but there would be a penetration of color into the, into this other stream, which is predominantly AB. this penetration of color is possible only if this SA, if the AB that I introduced is actually the AB corresponds to the sample antigen. But this penetration of color is not possible, if this AB that I introduced is not it does not correspond to the sample antigen. So, I have several such channels in place, I have several such and suppose, I can assume this is my sample and this sample can contain say ten such pathogens, it is possible. I want to find out which one is present there.


So, I have the target. So, I identify the target antigens and I get the antibodies and I put them in place and I flow them side by side; and I find, in one case there is diffusion of color deep into that other stream, deep into the antibody stream. And in other cases, the color remains mostly next the interface. So from that I can figure out which one is the target antigen. So, which pathogen I have in this sample. So, what we have here is that I said AB binds with SA, lot of free LA etcetera and spread of color into AB stream. More than one T-Sensor in a chip with different AB to target analyte, spread of color can be determined digitally. You can have an light source, you can have a light source and you can have a detection element placed on the next to the wall of the microchannel.

And from that, they can sense a digitally, it can go through some algorithm and it can tell you, this is the pathogen directly with though in that from that scanner. I mean, we do not have to look at this and find out how much the rate color as gone in and make decisions on your own, it is not that way. If it is it you can acquire the data digitally and have a software to analyze this and software to tell you this is the pathogen. So, this is device which can be used to find out which pathogen is there in a sample.

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H-filter

- Two parallel laminar streams will flow. One is the sample stream (e.g., blood containing aggregates of different sizes). The other is acceptor reagent (e.g., saline water).
- Smaller molecules diffuse faster.
- Smaller components of the sample stream will diffuse into the acceptor stream.
- At the end of the channel, two parallel flows are split up into two reservoirs.




There is another element of interest here, I mean which is unique these elements are unique. I mean, you we thought that we know about flow and macro scale processes, but in micro channel micro scale processes there are some simple elements that can do a lot of important work. Here you have two parallel lamina streams, two parallel lamina streams will flow; one is the sample stream, such as blood containing aggregates of different sizes and the other is acceptor reagents, such as saline water. So, what you have in this case is you have a channel through which you have two streams flowing and they are flowing parallel to each other, one is blood containing aggregates, blood containing aggregates of different sizes and the other one is saline water. So, you have these two streams flowing side by side. The smaller molecules, the smaller components, smaller particles in blood that you will find that will be very easy for these particles to diffuse into the water stream; whereas, larger ones the diffusion will be easy.

So, if you collect these two streams out, what do you have? You split the two flows into reservoirs. So, what you find there is the upper stream that will contain blood without the smaller particles or smaller aggregates, because smaller ones have gone to the saline water. So, if you want to separate the smaller ones from blood or if you want to concentrate the blood or focus on the bigger particles within the blood; so you can have such device. So, I said at the very outside that these laminar flow. And diffusion between two parallel parallelly flowing layers, these would be used in several elements of this micro scale device and they do a very unique and very useful job.

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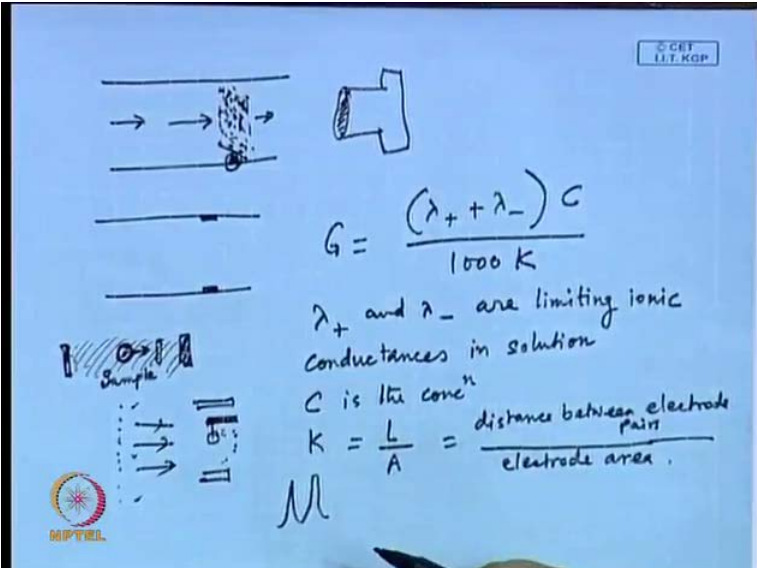
Detection

- **Laser induced fluorescence system**
 - Fluorophores are conjugated with migrating analytes
 - Laser beam excites the fluorophores
 - Resulting fluorescence signal is filtered to block background illumination from the excitation source.
 - Fluorescence signal is recorded using CCD camera, PMT, APD
- **Electrochemical**
 - Monitor variation of electrochemical potential as analytes migrate past a working electrode, positioned within the separation channel.
 - The conductivity is related to the concentration of species.



We discussed about this detection, we said there are two types of detections possible; one is a optical interrogation that is what we said, laser induced fluorescence system. And the other is the electrochemical method. In case of laser induced fluorescence system, fluorophores are conjugated with migrating analytes; laser beam excites the fluorophores, resulting fluorescence signal is filtered to block background illumination from the excitation source and fluorescence signal is recorded using CCD camera. What these means is that you intentionally put a fluorescing, you intentionally put some fluorophores, they are conjugated with migrating analytes.


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$$G = \frac{(\lambda_+ + \lambda_-) C}{1000 K}$$

λ_+ and λ_- are limiting ionic conductances in solution
 C is the concⁿ
 $K = \frac{L}{A} = \frac{\text{distance between electrode pair}}{\text{electrode area}}$



That means, I have a channel and I have some analytes flowing through this; and I am introducing fluorophores that are fluorophores conjugated with migrating analytes. What that means is, suppose I have say calcium ion and that calcium ion I want to identify. So, I introduce a fluorophore, which would be, I introduce a fluorescing agent that ties up with calcium. So, they are tied up with calcium and then what we do is we put a laser beam. We put a laser beam, so we put the beam here; and within this beam say I put a laser beam, I illuminate this layer by using a laser source using this light source; I illuminate this layer. And this is continuously this stream is flowing through this. So, I illuminate this layer, I illuminate this layer means, I produce a light of certain wavelength; that is what the purpose of laser is.

It will produce light of certain wavelength that will go and heat the fluorophores. And you get the light; you get a fluorescence signal, which is probably at a different wavelength. And that signal is captured by a photo detector, say for or simply a camera; suppose this is a camera, by a CCD camera, by PMT photo multiply tube or APD, which is avalanche photodiode. By some photo detector, you detect the light that is emanated that light that is coming out from this fluorophores. So, you intentionally add some chelating agent that ties up with the ion that you want to measure, say calcium ion you are measuring so it ties up with calcium.


And then, you find out what is the concept, then you introduce a light of certain wavelength and you know that the light, this light will when they when it hits this fluorophores, you will get a fluorescence signal of certain wavelength. Now on this camera or on this photo detector, if you have some kind of screen, some kind of filter to block the other the illuminations of other wavelengths; and only focus on that particular wavelength which this fluorophores can generate, then you can get a fair idea, what to what is exactly the concentration of the sample that you have. So, you have this resulting fluorescence signal is filtered to block background illumination from the excitation source, fluorescence signal is recorded using CCD camera PMT or APD. So, this is a laser induced fluorescence system, this particular source, this laser source and the detector, etcetera; these would be a part of the scanning device. So, these may not have to be a part of the microchannel, but this would be next to the microchannel and if this microchannel this microscale this wafer, if this is optically transparent, then it can be held the laser that that light source on the detector can be held next to it.

Now, there are other methods as well. For example, this electrochemical method is also there. In this method, what is written here is monitor variation of electrochemical properties, as analytes migrate past a working electrode, positioned within the separation channel, the conductivity is related to the concentration of species; that means, in the channel, you have electrodes positioned within the separation channel. So, you have electrodes positioned within the separation channel and you find out the conductivity as this quantity. Here this λ^+ , these and λ^- these are limiting ionic conductance limiting ionic conductances in solution; C is the concentration; K is the cell constant. K is the cell constant that is basically that L is distance between, so this is basically distance between electrode pairs divided by the electrode area. So by looking at this conductivity and by having a precalibration done, you can figure out which component is eluting from the channel at what time. So you can identify the components.

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Electrophoresis

- Migration behavior of charged species under the influence of an electric field.
- Analytes are suspended in an ionic buffer environment at a specific pH
- Each species migrates with a different mobility, allowing them to be resolved as distinct zones, and separated on the basis of size and charge.
- Biological macromolecules (e.g., proteins) are analytes
- Drag and electrophoretic mobility are the counteracting forces. Gravity is neglected.
- In some cases, polymer gel acts as sieving matrix material in the separation channel. The gel matrix reintroduces a size-dependence to the electrophoretic migration.
- In gel electrophoresis, analytes travel through the porous gel network with smaller fragments experiencing less resistance and eluting faster



I mentioned about this electrophoresis as one of the separation element. What we have in this electrophoresis here is that migration behavior of charge species under the influence of an electric field. Here analytes are suspended in an ionic buffer element at a specific pH. Each species migrates with a different mobility, allowing them to be resolved as distinct zones and separated on the basis of size and charge. Biological macromolecules are, biological macromolecules such as proteins are analytes. What we are talking about here is basically I have two electrodes. I have two electrodes and in between I have this

sample; so I have say one ion here, it has a size, it has a charge. And depending on what size and what charge it would be attracted towards certain electrodes. That is what we are talking about. Migration behavior of charge species under the influence of an electric field, analytes and suspended in an ionic buffer element at a specific pH; so the pH is at a, at this is done at a specific pH.

Each species migrates with a different mobility. Now these, see these particles would be experiencing what. These particle under this electric field, it would be pooled and at the same time, there would be a drag experienced by this particle. So depending on, so there would be a force balance, drag and electric phonative mobility they are counteracting forces and typically gravity is neglected. So each species migrates with a different mobility, allowing them to be resolved as distinct zones.

So, if you start this process and may be freeze after sometime, take away the electric field; what you will find is, these particles they have rearrange, they have repositioned themselves depending on their mobility. So, this is one way of say forming the or resolving the resolving this sample as into indistinct zones, separated on the basis of size and charge. So, you have distinct zones, suppose this sample contains a, b, c, d, e, f, g, h, out of them a, h, and k these belong to the same category, as far as the size and charge is concerned. So, they will band together in one location. Some other components, they will band together in another location. So, that is that would be eventually that is what is going to happen, if you have such system in place. And in some cases, on top of this, you have a polymer gel placed between these electrodes. So, basically sample is diffusing through this sample is moving through this polymer gel.

So in some cases, polymer gel acts as sieving matrix material in the separation channel. Gel matrix reintroduces a size dependence to the electrophoretic migration. That is in gel electrophoresis, analytes travel through the porous gel network with smaller fragments experiencing less resistance and eluting faster. Do you understand what a gel is, hydrogel it is basically made of water and small amount of polymer and cross linker is present there. So, this polymer chains they are cross link; that means, it could be possible that this cross linker that forms some kind of coordination complex between with the polymer. So, there are several dangling chain of the polymers and they are tied together by the cross linker. So, you form a network here and water remains trapped within this network; so that is typically a hydrogel. So that is a gel we see.

Now this so, it has a structure. So gel has a structure, because these chains are all ready cross linked; and water remains strapped within this cross linked network. So, this cross link network this acts as a sieving matrix; that means, if the molecule with a sample I said contains a, b, c, d, e, f, etcetera, out of them, the components that are small that can move through this network easily, where as the components which are big that cannot move through this network. So by so, you are in you are reintroducing, you are introducing another size dependence; you are introducing another size, another classification technique within the channel. So, this is basically the, so this is referred as gel electrophoresis. So, in gel electrophoresis analytes travel through the porous gel network with smaller fragments experiencing less resistance and eluting faster.

Now I want to, I want you to think of this that the picture that I had here, the picture that I have here, now if you invert this invert it this way, you have the electrodes here and the sample is placed in between. So, if you move it by 90 degree. And then you have a laminar flow through this in laminar flow in this direction. Let us do not confuse ourselves with the presence of gel, you have the two electrodes and then you orient them you rotate this by 90 degree. So depending on the size and the charge, the particles as I said a, d, and h, would be sitting in at one layer. So, if you have the two electrodes present there and if you have the sample, so they would be attracted to, I said it is moved by 90 degree; it is oriented, it is rotated by 90 degree. So, if after sometime if you switch off, if you freeze then this you will find one layer, one band with a, g, and h, depending on the size and charge; another band with some other components that has similar mobility.

So, you will be producing bands within, so within this channel. And now, if you have are slowing, if you are having this laminar flow, if you have a parabolic velocity distribution, one layer sliding against the other at different velocity. So, what you will find this you would be eluting layers based on their size, and charge. So, you will be producing as I said fractogram; one size, and then another size, and then another size. So, this is exactly what you call electrical field flow fractionation. I just now mentioned field flow fractionation of electrical type, where you have electric field to put the components in bands, and then eluting it using a parabolic velocity profile that is exactly what is electrical field flow fractionation. And that is what you can accomplish using electrophoresis in a rotated frame. I would like to stop here. In the next class, I will probably

complete this lab on a chip complete this discussion on lab on a chip, and then we will proceed further on basics of microfabrication. I stop here today.